

Help & Manual: -h | --help | --man | perldoc <cmd>

bioseq: Sequence Utility

FASTA descriptors

-l --length	Length of sequences
-n --num-seq	Number of sequences
-c --composition	Base or aa composition

FASTA filter - Multiple sequences

-r --revcom	Reverse-complement sequence
-p --pick 'tag:x'	Pick seq by tag ("id", "order", or "regex")
-d --delete 'tag:x'	Delete seq by tag ("id", "order", or "regex")
-t --translate 'n'	Translate in 1,3 or 6 reading frames
-g --no-gaps	Remove gaps

FASTA filter - Single sequence

-s --subseq 'x,y'	Sub-sequence from positions x to y (inclusive)
-R --reloop 'x'	re-circularize a bacterial genome at position x

Other options

-B --break	Write a FASTA file for each sequence
-C --count-codons	Count codons for sequence
-F --feat2fas	Extract FASTA sequence from GenBank bacterial genome file
-H --hydroB	Return Kyte-Doolittle hydropathicity (proteins)
-G --lead-gaps	Count and return leading gaps
-X --remove-stop	Remove stop codons
-x --restrict 'RE'	Predict fragments from a restriction enzyme digestion
--restrict-coord 'RE'	Predict fragments from restriction enzyme digestion in BED format
-o --output 'format'	Specify output file format. Default is "fasta". Optional format is "genbank"
-i --output 'format'	Specify Input file format. Default is "fasta". Optional format is "genbank"
-L --linearize	Linearize one sequence per line
--split-cdhit	Parse cdhit output .clstr file and generate a FASTA file for each CDHIT family

biotree: Tree Utility

-i --input 'format'	Specify Input file format
-l --length	Print total tree length
-m --mid-point	Midpoint root a tree
-u --otus-num	List all OTUs
-d --del-otus 'a,b,c'	Delete OTUs
--depth 'n1,n2,n3'	Print depth to root for nodes
--distance 'n1,n2'	Distance between two nodes
-D --del-low-boot'0.9'	Delete low-support (<0.9) branches
-r --reroot 'otu'	Reroot with "otu" as outgroup
-o --output 'format'	Output tree in "nhx" or "tabtree"
-c --ci 'trait-file'	Consistency indices for binary traits
-B --clean-boot	Remove branch support values
-b --clean-br	Remove branch lengths
--ead	Edge-length abundance distribution
--label-nodes	Append IDs to all nodes
--lca 'n1,n2,n3'	Return ID of the last common ancestor
-L --length-all	Print all nodes/branch length
-ltt 'number_of_bins'	Data from Lineage-through-time plot
--multi2bi	Multifurcating tree → bifurcating tree
-U --otus-desc 'n all'	Print all descendant OTUs of a node or all nodes
--random 'n'	Build tree of random subset of n OTUs
--sis-pairs	Print whether or not sisters for all pairs of OTUs
-s --subset 'otu1,otu2,otu3 innode'	Build tree for specified OTUs or a clade defined by an internal node
-t --as-text	Draw tree in ASCII text (for preview)
--tree-shape	Print input for R Package apTreeshape

-w | --walk 'out'

Print distances to all other OTUs from an OTU

biopop: PopGen Utility

-s --seg-sites	Print number of segregating sites
-p --pi	Print average pairwise nucleotide difference
-f --four-gametes	Perform four-gamete tests for each SNP pair
-c --snp-coding	Print SNP statistics for coding sequences
-C --snp-coding-long	Print the above in long format
-n --snp-noncoding	Print SNP statistics for coding or non-coding seqs
-m --mis-match	Output data for mis-match distribution
-b --bi-sites	Retain binary informative sites
-H --heterozygosity	Print heterozygosity for each SNP site
--bi-part	Print binary Newick trees for all SNPs
-b --bi-sites	Print alignment for binary-informative SNPs
--bi-sites-for-r	Print above to be read by R package "genetics"
-t --stats 'tag'	Statistics ('pi', 'theda', 'tajima_d', per-site values)

bioaln: Alignment Utility

Alignment descriptors

-l --length	Length of alignment
-L --list-ids	List sequence IDs
-n --num-seq	Number of aligned sequences
-a --avg-pid	Average percent identity
-w --window 'n'	Average difference by sliding window of size n.

Alignment viewers

-c --codon-view	Codon view (in groups of 3 nucleotides)
-m --match	Match view (highlight variable sites)

Alignment filters

-d --delete 's1,s2,s3'	Delete sequence(s)
-p --pick 's1,s2,s3'	Pick sequence(s)
-i --input 'format'	Specify input format. ClustalW is default.
-o --output 'format'	Specify output format. ClustalW is default.
-g --no-gaps	Remove gapped sites
-r --ref-seq 'seq_id'	Use seq_id as reference sequence
-s --slice 'x,y'	Return an alignment slice from x to y (inclusive)
-u --uniq	Remove redundant sequences
-v --var-sites	Show only variable sites
-P --pep2dna 'cds.fas'	Back align CDS to peptide alignment
-D --dna2pep	DNA alignment to protein alignment

Evolutionary analysis

-A --concat *.aln	Concatenate multiple alignments
-B --con-blocks 'n'	Extract conserved blocks of size n
-S --shuffle_sites	Make a column-permuted alignment
-R --resample 'n'	Resample n aligned sequences
-b --boot	Bootstrap an alignment
-M --permute-states	Permute within columns (to test tree-ness)
--remove-third	Remove third site
-I --aln-index 'id,n'	Return unaligned position for a sequence at n
--binary	Transform sequences into binary format
--bin-inform	Print only binary informative sites
-C --consensus 'n'	Add an n% consensus sequences
--gap-states, --gap-states2	Print gap statistics per column
-F --no-flat	Turns on 'begin-end' naming
--phy-nonint	Generate non-interleaved PHYLIP output
-E --rm-col 'id'	Remove columns with gap in sequence
--select-third	Generate alignment of every-third base
--trim-ends	Remove 5' and 3' gapped columns
--upper	Make uppercase alignment